Data Sheet

Olaquindox Metabolite(MQCA)ELISA Kit

Cat. #SG-4041

Size: 96 Wells

Principle and Application

This kit adopts the method of indirect competitive enzyme-linked immunoassay (ELISA) to detect Olaquindox Metabolite (MQCA) in the sample such as animal tissue, liver. The kit is composed of Microtiter Plate coated with coupled antigens, HRP conjugate, antibodies, standards and other supporting reagents. During the detection, with adding standards or samples, the MQCA in the samples will compete with the coupled antigens to combine with anti-MQCA antibodies. After adding HRP conjugate, take coloration with TMB substrates. Absorbance value of the samples is a negative correlation with MQCA content. Lastly, by comparing the obtained absorbance values with the standard curve, we can calculate the MQCA content in the sample.

Storage conditions

- The kit shall be stored at 2-8 ℃. Avoid freezing.
- Shelf life: 12 months. The date of manufacture is presented in the label of the box.

Technique Data

- Kit Sensitivity: 0.2ppb (ng/mL)
- Reactive Mode: 25℃, 30min～30min～15min
- Detection Limits:

Sample	Detection Limits
Tissue	0.4ppb
Liver	0.8ppb
Serum, urine	1 ppb

- Cross-reaction Rate:

Olaquindox Metabolite …………………………...……100%

Quinoxaline-2-Carboxylic Acid (QCA)……………….... < 1.0%

Olaquindox…………………………………………… < 1.0%

N-(2-hydroxyethyl)-3-methylquinoxaline-2-carboxamide...<1.0%

Quinocetone……………………………………………<1.0%

Desoxy-quinocetone (DQCT))…………………………<1.0%

Mequindox………………………………………………<1.0%

- Sample Recovery Rate:

Sample	Recovery Rate
Tissue, urine	85±25%
Liver, serum	80±25%

Composition of the Kit

Reagent	Specification
Microtiter Plate	8wells× 12strips
Standard: 0ppb, 0.2ppb, 0.6ppb, 1.8ppb, 5.4ppb, 16.2ppb	1.0mL each
High Standard (red cap)	100ppb
Antibody solution (blue cap)	1×5.5mL
HRP conjugate (red cap)	1×11mL
Substrate Reagent A (white cap)	1×6mL
Substrate Reagent B (black cap)	1×6mL
Stop Solution (yellow cap)	1×6mL
Concentrated Wash Buffer (20×)(white cap)	1×40mL
Concentrated Reconstitution Buffer(2×) (yellow cap)	1×50mL
Instruction	1
Adhesive Membrane	1
Sealed bag	1

Materials required but not supplied

- **Equipment**: microplate reader, printer, grinder (for homogenizing solid samples), nitrogen evaporator, vortex mixer (for shake and mix), centrifuge, graduated transfer pipette, and balance with a division value of 0.01 g, constant temperature device;

- **Micropipette**: single-channel (20-200μL and 100-1000μL), and multi-channel 300μL;

- **Reagents:** Ethyl acetate, concentrated sulfuric acid (mass fraction 98%), n-hexane.

Experimental preparation

Restore all reagents and samples to room temperature (adjust to around 25°C) for more than 30 min before use. This is a crucial step to ensure there is no precipitation in the reagents.

Please note that the labware must be clean. Use disposable pipette tips to avoid contamination of interference results.

◆ **Solution preparation:**

Solution 1: 2M Sulfuric Acid Solution

Add 10.9 mL of concentrated sulfuric acid slowly to deionized water, mix well, and make up to 100 mL. Store at room temperature.

Solution 2: Reconstitution Buffer

Dilute the Concentrated Reconstitution Buffer (2×) 2 times with deionized water (Reconstitution Buffer (2×): deionized water=1:1). It can be stored at 4 ℃ for one month.

Solution 3: Working Wash Buffer

Dilute the concentrated wash buffer (20×) by a factor of 20 (Concentrated wash buffer/Deionized water= 1: 19).

◆ **Sample pretreatment steps:**

1. Tissue (Pork, Chicken, Fish, Shrimp) treatment.

1) Weigh 1g ± 0.05g of defatted homogenized sample and add 8ml of **ethyl acetate** and 1ml of **2M sulfuric acid solution (Solution 1)**. Shake for 30 seconds and centrifuge at 4000 r/min at room temperature for 5 minutes.

2) Transfer 4mL of the upper ethyl acetate layer to a clean glass tube and evaporate it to dryness under nitrogen or air at 50°C-60°C.

3) Dissolve the dried residue in 1ml of **n-hexane**, shake for 30 seconds, then add 1ml of **reconstitution buffer (Solution 2)**, shake for 2 minutes, and centrifuge at 4000 r/min at room temperature for 5 minutes.

4) Remove the upper organic layer and take 50μL of the lower layer for analysis.

Dilution times of the sample:2   Detection limits: 0.4ppb

2. Liver treatment.

1) Weigh 1g ± 0.05g of defatted homogenized sample, add 8mL of **ethyl acetate** and 1mL of **2M sulfuric acid solution (Solution 1)**. Shake for 30 seconds and centrifuge at 4000 r/min at room temperature for 5 minutes.

2) Transfer 2mL of the upper ethyl acetate layer to a clean glass tube and evaporate it to dryness under nitrogen or air at 50°C-60°C.

3) Dissolve the dried residue in 2mL of n-hexane, shake for 30 seconds, then add 1mL of **reconstitution buffer (Solution 2)**, shake for 2 minutes, and centrifuge at 4000 r/min at room temperature for 5 minutes.

4) Remove the upper organic layer and take 50μL of the lower layer for analysis.

Dilution times of the sample:4   Detection limits: 0.8ppb

3. Serum treatment.

1) Take 1mL of serum (clear and non-hemolyzed) and add 4mL of **reconstitution buffer (Solution 2)**. Shake and mix for 2 minutes.

2) Take 50μL of the mixed solution for analysis.

Dilution times of the sample:5   Detection limits: 1ppb

4. Urine treatment.

1) Take 1mL of clear urine (centrifuge at 4000 r/min at room temperature for 10 minutes) and add 4mL of **reconstitution buffer (Solution 2)**. Vortex for 2 minutes.

2) Take 50µL of the mixed solution for analysis.

Dilution times of the sample:5   Detection limits: 1ppb

ELISA procedure

Place all reagents and samples to room temperature (adjust to around 25°C) for 30min. Gently shake the reagent bottles before use.

Take out the frame of the microplate along with the required number of wells. Then place the unused microplate wells into the sealed bag with the desiccant provided. Store the remaining kit in the refrigerator at 2-8°C.

Step 1: <u>Number</u>: Number the wells in sequence corresponding to the samples and standard, make 2-well parallel trials for each sample and standard, and record their locations.

Step 2: <u>Sample Incubation</u>: Add 50µL of **standard or sample** into each numbered well, then add 50µL of **antibody solution** into each well. Finally, cover the Microtiter Plate with the adhesive membrane, shake gently by hand (or use a microplate shaker) for 5s and incubate for 30 min at 25℃ in the dark.

Step 3: <u>Washing</u>: Uncover the adhesive membrane carefully, remove the liquid, pipette 350µL of **Working Wash Buffer (Solution 3)** to every well, let stand for 30 seconds then drain, repeat 5 times. Invert the plate and tap it against a thick absorbent paper (or lint-free cloth), with a soft towel placed underneath. (Bubbles that are not removed after tapping dry can be punctured with a clean pipette tip).

Step 4: <u>Enzyme Incubation</u>: Add 100µL of **HRP conjugate** into each well. Then cover the Microtiter Plate with the adhesive membrane, incubate for 30 min at 25℃ in the dark.

Step 5: <u>Washing</u>: Same as step 3.

Step 6: Color: Add 50μL of **Substrate Reagent A** to each well.　Then add 50μL of **Substrate Reagent B** per well. Shake gently by hand (or use a microplate shaker) for 5s, and allow to react for 15min at 25℃ in the dark. (The reaction can be extended appropriately if the blue color is too pale.)

Step 7: Stop the reaction: Pipette 50μL of **Stop Solution** to each well, and shake gently by hand (or use a microplate shaker). The reaction would be stopped.

Step 8: Calculate: Determine the Optical Density (OD value; absorbance value) at 450nm (Reference wavelength 630nm) with a microplate reader. Finish this step within 10min after stop the reaction.

Interpretation of result

◆ **Calculate the percentage of absorbance value**

$$\text{Percentage of absorbance value}(\%) = \frac{A}{A0} \times 100\%$$

A—the average OD value of the sample or the standard solution;

A0—the average OD value of the 0ppb standard solution).

It is used to calculate the percentage absorbance of a standard or sample.

◆ **Draw the standard curve and calculate**

- Take absorbance percentage(A/A0) of standards as Y-axis and the corresponding log of standards concentration (ppb) as X-axis.

- Draw the standard semi-log curves with X-axis and Y-axis.

- Take absorbance percentage of samples substitute into standard curve, then can get the corresponding concentration from standard curve. **Last, the resulting concentration values multiplied by the corresponding dilution times is the actual concentration of MQCA of samples.**

If professional analysis software of the kit is used for calculation, it is more convenient for accurate and rapid analysis of a large number of samples.

Attention

- Before test, the reagents and samples should be balanced to room temperature (25℃). If below 25℃, it will lead to all the standard OD value on the low side.
- In washing process, dry wells may result in non-linear standard curves and undesirable reproducibility. Therefore, proceed to the next step immediately after washing.
- Please mix the contents within the wells uniformly and wash the plate thoroughly. The reproducibility is largely determined by consistency of washing step.
- During the incubation, cover microplates with adhesive membrane to avoid light.
- Do not use kits that are overdue. Do not mix reagents with those from other lots.
- Substrate Reagent A/B is colorless. If not, please discard.
- If absorbance value of 0ppb is below 0.5（A450nm< 0.5 ）, it means that the reagent may be metamorphic.
- Stop solution is corrosives. Please avoid contacting with skin.
- **As the OD values of the standard curve may vary according to the conditions of actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.**
- **For the mentioned sample, fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
- The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS can be used for quantitative confirmation.